

Best Practice Sample Submission

Nasal/ Laryngeal / Bronchial / Joint Swabs



Sample collection

- Samples should be collected on a synthetic swab (Dacron or Rayon) with a plastic shaft
 - Do not use wooden shaft swabs (the wood will absorb the testing fluid)
- Submit swab in **Transport Media** (for Bacteriology) or **PBS**(For Molecular testing) - do not submit dry swabs. This does not apply to cheek swab submissions, see details below for specific cheek swab submissions.
 - **Transport Media**
 - Place swab in a sterile tube with liquid media-saturated foam (such as liquid Stuart or Hank's media)
 - Do not use agar gel-type media as this may inhibit PCR
 - Do not use media with additives (such as charcoal, antibiotics, or heparin) as this may inhibit PCR
 - Check that the swab cap is securely twisted shut
 - **PBS** (Phosphate Buffered Saline) **For molecular testing only:**
 - Place swab in a snap cap tube (1.5 ml or 5 ml) with 1 ml of 1X PBS
 - Trim swab shaft short enough for the lid of the snap cap tube to close securely
 - Check that the cap is securely snapped shut (5 ml tube must be double snapped)
- For ideal sample volume, submit 1 swab per test. If multiple tests are requested on a single swab, testing will be at the discretion of the pathologist, in order to accommodate available sample volume.

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- Label each tube with its swab type (Nasal, Laryngeal, Bronchial, Tracheal, Brain, Joint, etc)
- Label tubes consecutively (ex: Tube #1-10, or #21-60).
 - For 1.5ml tubes: number the top of the tubes
 - For all other tubes: number the side of the tubes
- Record individual Animal IDs on the submission form next to the corresponding Tube #.
- Do not include Animal IDs on the tube label as this can cause confusion. Following these labeling instructions will greatly expedite sample processing.